



Superoxide Anion Microplate Assay Kit User Manual

Catalog # ASK1171

Detection and Quantification of Superoxide Anion (SOA) Content in Serum, Plasma, Tissue extracts, Cell lysate, Cell culture media, Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.

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I. INTRODUCTION

Superoxide anion (O_2^-) is a short-lived radical of molecular oxygen that plays key roles in the immune system and intracellular functions. The enzyme nicotinamide adenine dinucleotide phosphate (NADPH) oxidase-acytochrome-b-558-containing, plasma-membrane-bound enzyme complex-synthesizes superoxide anion by transferring an electron to molecular oxygen. Superoxide anion is a potent oxidant, which is released by leukocytes to damage infectious organisms. Superoxide anion is also implicated in oxidative stress damage, tumor promotion, and cell growth and DNA synthesis, which superoxide anion affects through the cell signaling pathway of Ras.

Superoxide Anion Microplate Assay Kit is a sensitive assay for determining Superoxide Anion content in various samples. The color intensity at 530 nm is linear to the Superoxide Anion content in the sample.



II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Substrate	Powder x 1	4 °C
Dye Reagent I	Powder x 1	4 °C
Dye Reagent II	Powder x 1	4 °C
Standard	Powder x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

Note:

Substrate: add 2 ml distilled water to dissolve before use.

Dye Reagent I : add 8 ml distilled water to dissolve before use.

Dye Reagent II: add 8 ml absolute alcohol to dissolve before use.

Standard: add 1 ml distilled water to dissolve before use; then add 10 µl into 990 µl distilled water. The concentration will be 1 mmol/L.

III. MATERIALS REQUIRED BUT NOT PROVIDED

1. Microplate reader to read absorbance at 530 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Centrifuge
7. Timer
8. Absolute alcohol



IV. SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for 5×10^6 cell or bacteria, sonicate (with power 20%, sonication 3s, interval 10s, repeat 30 times); centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For tissue samples

Weigh 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

3. For serum or plasma samples

Detect directly.

**V. ASSAY PROCEDURE**

Add following reagents in the microplate:

Reagent	Sample	Standard	Blank
Sample	20 μ l	--	--
Standard	--	40 μ l	--
Distilled water	--	--	40 μ l
Substrate	20 μ l	--	--
Mix, cover the plate and incubate at 37 °C for 20 minutes.			
Dye Reagent I	80 μ l	80 μ l	80 μ l
Dye Reagent II	80 μ l	80 μ l	80 μ l
Mix, cover the plate and incubate at 37 °C for 20 minutes, record absorbance measured at 530 nm.			

Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) The concentrations can vary over a wide range depending on the different samples. For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.

VI. CALCULATION

1. According to the volume of sample

$$\text{SOA } (\mu\text{mol/ml}) = (\text{CStandard} \times \text{VStandard}) \times (\text{ODSample} - \text{ODBlank}) / (\text{ODStandard} - \text{ODBlank}) / \text{VSample}$$

$$= 2 \times (\text{ODSample} - \text{ODBlank}) / (\text{ODStandard} - \text{ODBlank})$$

2. According to the protein concentration of sample

$$\text{SOA } (\mu\text{mol/mg}) = (\text{CStandard} \times \text{VStandard}) \times (\text{ODSample} - \text{ODBlank}) / (\text{ODStandard} - \text{ODBlank}) / (\text{VSample} \times \text{CProtein})$$

$$= 2 \times (\text{ODSample} - \text{ODBlank}) / (\text{ODStandard} - \text{ODBlank}) / \text{CProtein}$$

3. According to the weight of sample

$$\text{SOA } (\mu\text{mol/g}) = (\text{CStandard} \times \text{VStandard}) \times (\text{ODSample} - \text{ODBlank}) / (\text{ODStandard} - \text{ODBlank}) / (\text{VSample} \times \text{W} / \text{VAssay})$$

$$= 2 \times (\text{ODSample} - \text{ODBlank}) / (\text{ODStandard} - \text{ODBlank}) / \text{W}$$

4. According to the quantity of cell or bacteria

$$\text{SOA } (\mu\text{mol}/10^4) = (\text{CStandard} \times \text{VStandard}) \times (\text{ODSample} - \text{ODBlank}) / (\text{ODStandard} - \text{ODBlank}) / (\text{N} \times \text{VSample} / \text{VAssay})$$

$$= 2 \times (\text{ODSample} - \text{ODBlank}) / (\text{ODStandard} - \text{ODBlank}) / \text{N}$$

CStandard: the standard concentration, 1 mmol/L = 1 $\mu\text{mol/ml}$;

CProtein: the protein concentration, mg/ml;

W: the weight of sample, g;

VStandard: the volume of standard, 40 μl = 0.04 ml;

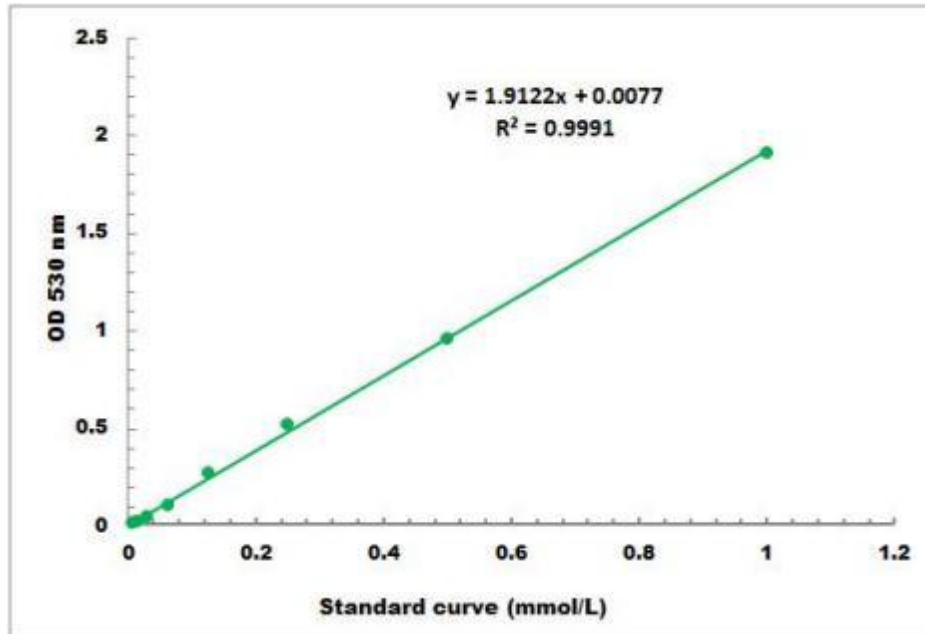
VSample: the volume of sample, 20 μl = 0.02 ml;

N : the quantity of cell or bacteria, $\text{N} \times 10^4$;

V_{Assay}: the volume of Assay buffer, 1 ml;

VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.01 mmol/L - 1 mmol/L